



Role of *Helicobacter*, An Emerging Zoonotic Pathogen in Gastro-intestinal Diseases

Gulzar Ahmad^{1*}, Zainab Zaki², Farha², Sunaiba Manzar², MD Moiz Alam²



¹Department of Veterinary Public Health and Epidemiology, IVRI, Bareilly, UP, India

^{2,3}Department of Ilmul Saidla, A.K.T.C.H, AMU, Aligarh, UP, India

ARTICLE INFO

Article History

Received 17 September 2020

Revised 8 March 2021

Accepted 9 March 2021

Available Online 11 March 2021

Keywords:

Helicobacter,

Zoonotic,

MALT lymphoma,

Gastroenteritis,

Unani drugs

ABSTRACT

Helicobacter is the organism widely spread in human and all known *Helicobacters* live in human and animal hosts especially *H. pylori*, where colonization occurs principally in the gastrointestinal tract and sometime responsible for serious illness such as gastritis, Peptic Ulcer Disease (PUD) and strongly associated with gastric carcinoma, MALT lymphoma. It is considered as a serious problem impairing in public health in both developing and developed countries because it colonises the gastric mucosa of about half of the world population. Although less often Non *Helicobacter Pylori Helicobacter* (NHPH) are also able to cause disease in humans. *Helicobacter* naturally infects many poultry birds, some rodent species as well as humans. *Helicobacter* is a zoonotic bacterium that has also been associated with certain enteric infection in humans. *Helicobacter* is of zoonotic importance and the animal host remains a natural reservoir for many species. And efforts for *Helicobacter's* treatment is more difficult due to antibiotic resistance and patient compliance but there are various unani drugs of plants, animals and mineral origin which are being used for the treatment of *Helicobacter's* infection. The aim of this review is to give a comprehensive knowledge of zoonotic potential of *Helicobacters*.

1. INTRODUCTION

The *Helicobacter* genuses are gram negative bacteria which were originally considered to belong to the *Campylobacter* genus because of resemblance in morphological characters. But, since 1989 they have been classified in a separate genus because of having different biochemical characteristics, more than 35 species having been identified till now and more still being studied (Fox *et al.*, 2002; Hua *et al.*, 1999). In this genus, the best known species is *H. pylori*, a specific pathogen of the upper gastrointestinal tract. *Helicobacter pylori* infection is one of the most common infections and nearly about half of the world population is suffering from this carcinogenic organism (Parsonnet *et al.*, 1999). It has been reported that *H. pylori* is almost always acquired in childhood (Rowland, 2000). It is the main cause of gasterointestinal diseases like gastritis, peptic ulcers, and gastric carcinoma. In 2005, Barry Marshall and Robin

Warren were awarded with the Nobel Prize in Physiology or Medicine "for their discovery of the bacterium *Helicobacter pylori* and its role in gastritis and peptic ulcer disease". The discovery of *H. pylori* increased interest to other spiral bacteria that had been seen in many animal species. Most of these bacteria belong to the genus *Helicobacter* (O'Rourke *et al.*, 2001). Other than *H. pylori* that is Non-*pylori helicobacters* (NPHS) are increasingly being found in human clinical specimens. Thus, their role in human medicine is increasingly reported.

Enterohepatic *Helicobacters* (EHH) other than *H. pylori* colonize the bowel, biliary tree and liver of animals and human beings with pathogenic potential (Bohr *et al.*, 2007) However, although the gastric *Helicobacters* have been the most studied, they only account for one third of the entire genus. The remaining two-thirds correspond to the so-called enterohepatic *Helicobacter* (EHH) (Schauer *et al.*, 2001).

The organisms are found about 70–90% and 25–50% of the population in developing and developed countries, respectively (Guillermo *et al.*, 2004; Sykora *et al.*, 2006; Vale and Vitor, 2010). This pathogen is reported as a cohabitant of the host (Cremonini and Gasbarrini, 2003), and it does not always cause illness in infected people and remain

*Corresponding Author: Gulzar Ahmad

E-mail Address: zzakiahmad15@gmail.com

DOI: 10.46890/SL.2021.v02i02.003

© 2021 by the authors. The license of Science Letters. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

asymptomatic (Santos *et al.*, 2009). Several risk factors for *H. pylori* infection have been known and this include poor social and economic development (Lehours and Yilmaz, 2007), low education level, poor hygienic conditions such as poor hygiene practices during childhood, lack of a household bath, absence of sanitary drinking water, absence of a sewage disposal facility during childhood, crowded families (Nouriaie *et al.*, 2009), and unhygienic food handling (Van Duynhoven and de Jonge, 2001). Most of these factors are a consequence of and associated with socioeconomic development. The prevalence of the infection decreases by the improvement of general hygienic conditions (Fujimoto *et al.*, 2007). The transmission pathways of *H. pylori* remain unclear. The most commonly accepted hypothesis is that infection can take place through fecal-oral route (Vale and Vitor, 2010) and contaminated food and water may play an important role in transmission of this organism to humans (Van Duynhoven and de Jonge, 2001; Gomes and De Martinis, 2004; Vale and Vitro, 2010). Indeed, *H. pylori* have been detected in drinking water (Queralt *et al.*, 2005) and foods of animal origin such as milk, and meat (Dore *et al.*, 2001; Fujimura *et al.*, 2002; Quaglia *et al.*, 2008). Due to that, the existence of animal reservoirs of this organism has been hypothesized (Dore *et al.*, 2001; Fujimura *et al.*, 2002). This hypothesis is further more supported by the demonstration of *H. pylori* in the gastric mucosa of calves, pigs, and horses and its isolation from sheep's gastric tissue and milk (Dore *et al.*, 2001), by this it can be suggesting that these animal species may act as reservoirs and are the spreaders of *H. pylori*. *H. pylori* can survive for long period in foodstuffs such as milk and instant foods (Poms and Tatini, 2001; Quaglia *et al.*, 2007). Therefore, food may serve as a vehicle for *H. pylori* infection.

Etiology

H. pylori are a helical or curved shaped Gram-negative bacterium and the name *Helicobacter pylori* is Latin for spiral rod of the lower part of the stomach. It has two to six flagella which varies in different species of *Helicobacters* and they helps in mobility to withstand rhythmic gastric contractions and penetrate the gastric mucosa. The primary reservoir for gastric *Helicobacters* like *H. pylori* (human stomach), *H. canis* (canine stomach), *H. suis* (pig stomach) and in case of NHPH (non-helicobacter pylori helicobacter) the site are others such as *H. pullorum* (poultry intestine), *H. hepaticus* (liver). All gastric *Helicobacters* contains a large amount of urease enzyme that produces urease, which makes the environment alkaline that enables to survive the the bacteria in acidic medium of stomach. On the other hand NHPH does not produce urease enzyme therefore they cannot create alkaline medium in stomach hence do not survive in stomach. *Helicobacters* have number of virulence factors, including VacA and CagA that may have different disease associations (Atherton, 1997). The optimum growth temperature is 35-37°C but some species of *Helicobacters* grow poorly at 42°C and 30°C. *Helicobacter* species are micro-aerophilic and grow best in an atmosphere of 86% N₂, 4% O₂ with 5% CO₂ and 5% H₂ (Goodwin, 1989). All the required growth conditions are met in the gastrointestinal tract of all warm-blooded animals. At temperatures below 30°C, *H. pylori* could survive in some foods, such as fresh fruit and vegetables, fresh poultry or fish, fresh meats, and some dairy products, water and milk (Fan *et al.*, 1998).

Routes of Transmission

Numerous epidemiological studies have been carried out to determine the factors affecting of this pathogen's transmission. Socio-economic status is obviously the most significant determinant for the occurrence of *Helicobacteriosis* infection, with a significantly higher prevalence of poorer/lower social group (Mitchell, 2001). These factors encompasses conditions such as levels of hygiene, density of living, sanitation, and educational opportunities, which have all been individually identified as markers of the bacterium presence.

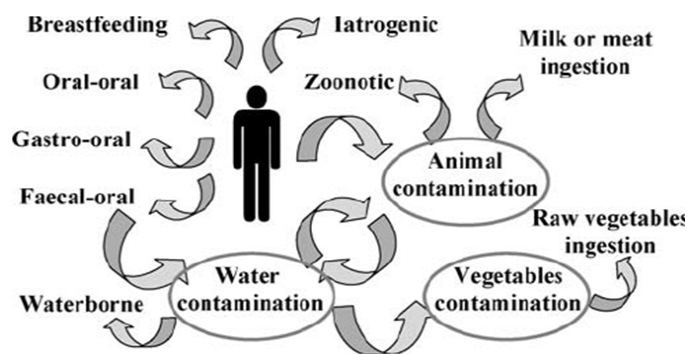


Fig. 1 Suggested transmission routes for *Helicobacters*. Five of the proposed pathways are representative of direct person-to-person transmission (breastfeeding, iatrogenic, oral-oral, gastro-oral, and fecal-oral), whereas the remaining four require an environmental reservoir in between. Possible reservoirs outside the human host are marked with a red circle.

On the basis of epidemiological and microbiological evidence, several routes of transmission have been conjectured as shown in above picture. Transmission from one person to another is widely viewed as the most likely route of infection. In addition, several epidemiological studies have consistently identified domestic overcrowding and infection of family members as a risk factor for its transmission. The most relevant pathways of person-to-person transmission include the gastro-oral, oral-oral, and fecal-oral routes. Iatrogenic and breastfeeding transmission are often used as possible means of transmitting the pathogen. Additionally, there are at least three potential vectors suggested to maintain the bacterium in viable form: water, food, and animals. Most scholar accept the relative importance of these routes in the transmission of the bacterium is likely to vary between developing and developed countries (Perez-Perez, Rothenbacher, and Brenner 2004; Megraud, 2003).

Zoonotic Transmission

An obvious reasoning, including contact with animals as a possible mode of transmission mode, since zoonotic transmission represents one of the leading causes of disease and death from infectious disease worldwide. Many epidemiological studies seem to support the role of animals in the acquisition of *H. pylori*, but the extent of this aid depends on the animals under study. Considered vectors include sheep (Dore *et al.* 2001), cows (Fujimura *et al.* 2002), domestic pets (Boomkens *et al.* 2004), houseflies (Osato *et al.* 1998) and cockroaches (Imamura *et al.*

2003). Epidemiological data showed a greater prevalence in shepherds and their families compared to the general population (Dore et al. 1999; Plonka et al. 2006). An epidemiological study has been shown controversial results in respect to the risk of the presence of domestic animals in the household (e.g., Bode et al. 1998; Lindo et al. 1999; Kearney and Crump 2002). *H. pylori* has not been found in dogs and only very rarely in cats' stomachs (ElZaatar et al. 1997; Neiger and Simpson 2000), and it has been suggested that the presence in animals is of human origin (Cittelly et al. 2002; ElZaatar et al. 1997). Recent studies have identified *H. pylori* by PCR in the bile of cats, thus increasing the chance of this animal being a vector (Boomkens et al. 2004). Nearly every animal is now considered to colonize by its own endogenous *Helicobacter* spp. As *H. pylori*, that has co-evolved with humans to be highly specialized in the colonization of the human GI tract (Falush et al. 2003), such bacteria have specialized in colonizing their specific natural host's GI tract. In the model where mammal's stomach is colonized by only one strain, *H. pylori* would find tough competition by these other *Helicobacter* spp. in search for essential nutrients and not subsist. With the emergence of a multiple infecting strains and species model for the same host it is more credible that *H. pylori* is a zoonotic agent as well.

Diseases associated with *Helicobacters*

Helicobacter pylori are the main cause of gastric inflammation in nearly all infected subjects (Farinha and Gascoyne, 2005). Most infected people show little to no symptoms, and only a small percentage will develop severe gastric disease (Suerbaum and Josenhans, 2007; Amieva and El-Omar, 2008). The pattern and distribution of chronic gastric inflammation is associated with the type of lesions found. Individuals with body-predominant gastritis and normal or decreased acid production are more likely to develop gastric ulcers, gastric atrophy, gastric intestinal metaplasia and eventually, gastric carcinoma while individuals with antral-predominant gastritis, which is the most common form, show increase acid production and increased risk to develop duodenal ulcers (Kusters et al., 2006; Lochhead and El-Omar, 2007).

The next phase of human *H. pylori* infection is regulated by changes of the epithelial cell cycle, particularly increased rates of apoptosis and cell proliferation. These alteration may be responsible for the multifocal atrophy which characterizes the type of gastritis associated with the increased risk of cancer. During this advanced phase nuclear and architectural abnormalities become apparent, which can reflect progressive mutational events as expected in classical molecular models of carcinogenesis (Correa, 2004). In spite of hereditary and other known factors *H. pylori* infection is one of the major risk factor for developing cancer in human (Gonzalez et al., 2012; Wadhwa et al., 2013). Nearly about 1–3% of people infected with *H. pylori* develop gastric cancer. Even though only a small proportion of patients with *H. pylori* will ultimately develop malignant disease, the widespread high prevalence of this bacterium explains that gastric cancer remains the fifth most common cancer in the world (Globocan, 2012). It is also documented as the third common cause of cancer-related death in men and fifth in women (Torre et al., 2015; Chmiela et al., 2017). In 1994 the International Agency for Research on Cancer classified *H. pylori* as a class I carcinogen on the basis of

epidemiological evidences (IARC, 1994; Chmiela et al., 2017). In fact, *H. pylori* infection plays a key role in the development of two different gastric malignancies: gastric adenocarcinoma and gastric MALT lymphoma (Parsonnet, 1994; Stolte et al., 2002; Farinha and Gascoyne, 2005; Suerbaum and Josenhans, 2007; Patel et al., 2014).

Nearly 60% of the intestinal type gastric cancer is due to *H. pylori* infection. Chronic *H. pylori* infection causes genetic instability in gastric epithelial cells and affects the DNA damage repair systems. Consequently, *H. pylori* infection should always be treated as pro-cancerous factor (Chmiela et al., 2017). In 1991, The association between MALT-lymphoma and *H. pylori* was firstly reported in 1991 (Wotherspoon et al., 1991) and it is responsible for 92 to 98% of gastric MALT-lymphomas (Mbulaiteye et al., 2009) throughout the world.

The occurrence of NHPH infection in human patients is lower in comparison to *H. pylori*, but it is probably an underestimation of the real infections because NHPH screening is not frequent (Kawakubo et al., 2018). Until now, 5 gastric NHPH species have been reported in human patients suffering from gastric disorders. In Belgium, Germany and China, it has been revealed that *H. suis* is the most common NHPH species in human patients suffering from gastric disorders, followed by *H. salomonis*, *H. felis*, *H. heilmannii* and *H. bizzozeronii*. (Van den Bulck et al., 2005; Liu et al., 2014). Although less commonly, gastric NHPH may also be able to cause disease in humans. NHPH infections in human stomach may be accompanied by acute gastritis (Lavelle et al., 1994; Yoshimura et al., 2002), active chronic gastritis (Haesebrouck et al., 2009), erosions mainly located in the antrum (Debongnie et al., 1998; Seo et al., 2003) and duodenal ulcers (Jhala et al., 1999; Iwanczak et al., 2012). Moreover, some species (*H. mustelae*, *H. hepaticus* and *H. bilis*) exhibit carcinogenic potential in animals and harbor several virulence genes and may cause diseases not only in animals, but also in humans. The presence of NHPH was significantly associated with large intestinal carcinoma (68%) and mucinous adeno-carcinoma (92%) in man and animal (Swennes et al., 2016). Glandular atrophy and intestinal metaplasia of the fundic gastric mucosa were also reported. Still, these lesions appear to be less common and less severe than those which are associated with *H. pylori* (Yoshimura et al., 2002). Infection of Human NHPH such as *H. heilmannii*, have also been associated with low-grade MALT lymphoma of the stomach as well as other type of gastric cancer and the risk of developing this disease is higher for NHPH than *H. pylori* (Haesebrouck et al. 2009; Joosten et al., 2015). They have been reported to determine after clearance of the NHPH, therefore, emphasizing a causal relationship (Morgner et al., 2000; Thomas-Marques et al., 2005). Many human patients infected with NHPH species are asymptomatic (Mazzucchelli et al., 1993) while others have atypical complaints like acute or chronic epigastric pain and nausea, hematemesis, recurrent dyspepsia, irregular defecation frequency and consistency, vomiting, heartburn, dysphagia and loss of appetite (Dieterich et al., 1998; Mention et al., 1999; Kaklikaya et al., 2002; Seo et al., 2003). Most human isolates are cultured rarely *in vitro*; therefore NHPHs associated with human chronic gastritis have been poorly characterized (Kawakubo et al., 2018). Thus, the clinical outcome of *Helicobacter* infection is diverse and dependent on the strain, virulence characteristics, host genetic susceptibility, environmental factors and their interactions. *Helicobacter*-like organisms are often found in canine stomachs including healthy

dogs and those showing signs of gastrointestinal disease, but the relationship between such organisms and gastric pathology has not been well-known. Some of such organisms have zoonotic importance (Asl et al., 2010). The predominant gastric NHPH spp. found in dogs are *H. bizzozeronii*, *H. felis* and *H. heilmannii* (s.s.), whereas *H. salomonis* has not often been detected. *H. cynogastricus* was recently isolated from the stomach of a dog (Haesebrouck et al., 2009). *Helicobacter canis* has been associated with hepatobiliary and gastrointestinal disease in dogs, cats and humans (Hristova et al., 2017). Though less frequent, the role of *H. canis* in the etiology of gastric cancer was also observed where in a recent Norwegian survey, canine GC accounts for 0.16% of all reported canine cancer cases (Seim-Wikse et al., 2013). Similarly, *Helicobacter suis* mainly colonizes from the porcine stomach and seems to be the most important among the NHPH. It is mostly associated with gastric pathologies in animals (Haesebrouck et al., 2009) and also the most commonly detected NHPH species in humans (De Groote et al., 2005; Van den Bulck et al., 2005; Liu et al., 2014) where it may cause gastritis, peptic ulcer disease and gastric mucosa-associated lymphoid tissue (MALT) lymphoma (Flahou et al., 2012; Matsui et al., 2014). Infections with this bacterium have also been associated with decreased daily weight gain in pigs from 5 to 10 % (Haesebrouck et al., 2009; Kumar et al., 2010). Pigs are the main reservoir of *H. suis* (De Bruyne et al., 2012) Therefore, this organism has also been found in the stomach of rhesus and crab-eating macaques (Bosschem et al., 2016). In human, histologic analysis of biopsies from a patient with asymptomatic nodular gastritis also discovered the presence of spiral-shaped bacteria in the gastric mucosa, indicative for a NHPH infection. Besides this, it has also been detected in 27% of patients with Parkinson's disease (Blaecher et al., 2013 and 2017). Likewise, the presence of co-infections in human patients and an increased prevalence of peptic ulceration during co-infection have been reported (Liu et al., 2014; Overby et al., 2017).

H. pullorum infection has been linked to vibronic hepatitis and enteritis in chickens (Stanely et al., 1994). It is also associated with gastroenteritis and hepatobiliary disease in humans (Ceelen et al., 2006; Turk et al., 2012). Moreover, *H. pullorum* has been detected in humans with gastroenteritis, women suffering from chronic cholecystitis and a patients suffering from cirrhosis and/or hepato-cellular carcinoma. The species reportedly has also been isolated from a diarrhetic psittacine bird, suggesting that pet birds maybe a zoonotic risk for humans (Ceelan et al., 2006; Turk et al., 2012).

Diagnosis

For clinical purposes, several methods can be used to diagnose *Helicobacter* infection, which are divided into two main methods that is invasive and non-invasive methods which are based on the use of endoscopy (Kamboj et al., 2017). Histopathology, culture and rapid urease test (RUT) which is widely available and employed worldwide are considered as the invasive tests. Fecal antigen test, urea breathe test (UBT) and serology are known as the non-invasive tests (Guarner et al., 2010; McColl et al., 2010).

Invasive tests

Invasive methods are endoscopic biopsy based procedures and include brush cytology, urease test, histological examination, electron microscopy, culture, the polymerase chain reaction techniques (PCR). There has been a suggestion that some

factors capable of affecting the performance of all biopsy-based tests, such as site, number and dimension of biopsies or a previous eradication attempt, which can significantly decrease the diagnostic accuracy. Culture methods have the advantages of allowing antimicrobial susceptibility testing and detailed characterization of the cultured organism. However, the culture of *H. pylori* from gastric biopsy specimen is the gold standard methods for diagnosis of infection, the culture of this bacterium typically take 3-5 days and hence are very slow. Endoscopy samples can also be assessed by histological methods. *H. pylori* can be visualized with hematoxylin and eosin (H&E) staining of tissue sections. There is a possibility of false identification with this stain. Hence, a sensitive staining technique consisting of a combination of H&E staining, Steiner silver staining and alcian blue staining is used for more accurate results (Genta et al., 1994). Another application of endoscopy samples is in the tissue urease tests. However, these tests are dependent on the bacterial load in the stomach. Biopsy samples obtained by endoscopy can also be used for PCR analysis.

Non-invasive tests

As the invasive procedure are expensive, unpleasant to patients and carries a small but definite risk of complications the non-endoscopic tests to assess *Helicobacters* infection have been welcomed. The more widely available non-invasive procedures for diagnosing *Helicobacters* infection are the urea breath test, serological assays and detection of bacteria, as well as bacterial DNA and antigens in stool. A recently developed stool assay has been shown to be a reliable diagnostic tool and is becoming increasingly available and employed.

Unani approach for the treatment of *Helicobacter*

WHO has classified *Helicobacter pylori* as a class I carcinogen and the eradication of this silent killer with antibiotics (triple therapy) combinations has been reported to be beneficial in preventing gastric ailments especially cancer (IARC, 1994). However, increasing dilemma of antibiotic resistance, adverse effects and high costing has lead researchers to explore natural resources especially plant materials as an alternative source of antimicrobials.

Unani physicians listed a broad range of care for this disease based on the causes, clinical presentations, locations, environments, age, acuteness or chronicity, and dietary habits with primary concern for the correction of patients' Mizaj (temperament) and Akhlat (humours).

In Unani, medicinal plants, animals as well as drugs of mineral origin are being used for the treatment of chronic gastritis which is mainly due to *H. pylori* without any reported side effects e.g some of widely used single drugs (adviya mufrida) are *Aloe barbedensis* Mill (*Elva*), *Alpinia galangal* Willd (khulanjan), *Althaea rosea* Linn (khatami), *Anchusa strigosa* Labill (Gaozaban), *Glycyrrhiza glabra* Linn (Asl-us-soos), *Withania somnifera* Linn (Asgandh), *Andrographis paniculata* Wall (Bhuineem), *Zingiber officinale* Rosc (Adrak), *Picrorhiza kurroa* Royle (Kutki), *Emblica officinale* (Amla), *Nigella sativa* Linn (kalonji), *Momordica charantia* Linn (Karela), *Curcuma longa* Linn (Haldi), *Asparagus racemosus* Willd (satawar), *Aegle marmelos* Correa (Bael), *Myristica fragrans* Houtt (jaiphal) etc and in the form of compound drugs (Adviya murakkaba) are Majoon Dabidul Ward, Jawarish Anarain, Sharbat Anar, Majoon Zanjbil,

Jawarish Mastagi, Qurs Satawari, Itrifal Aftimoon, Sharbat Unnab and Khameera Sandal have been indicated by Unani physicians for the treatment of chronic gastritis (*H. pylori*) and their efficacy against gastritis has also been tested by in vivo and in viro studies. (Aziz ., Jurjani., al., Qurrah., Marwan., Mohd et al., 2011). Those drugs mode of action is both systemic and local. Such medicines, apart from the correction of altered mizaj (temperament), have a claiming effect on the inflamed mucosa, provide ground material for healing, eliminate inflammatory factors and also have antiseptic or antibacterial impact. In spite of that physicians have recommended different seasons, regions and mizaj. Some researchers demonstrate the effect of Unani compound drug containing “ *Glycyrrhiza glabra* (*Asl-us-soos*), *Plantago ovate* (*Aspghol musallam*), *Acacia arabica* (*samagh-arabi*) and *Pistacia lenticus* (*Mastagi*)” for the treatment of gastric anomalies and eradicating *H. pylori* bacteria from the stomach (Rahida et al., 2017). *Asl-us-soos* (*Glycyrrhiza glabra*)/ *licorice* reduces stomach secretion and produces thick mucus that protects the lining of stomach from inflammation, gastritis and peptic ulceration, it contains flavonoids, therefore an anti-inflammatory and anti-bacterial effect (Khare et al., 2004; Kakka et al., 1998) *Mastagi* (*Pistacia lenticus*) is cytoprotective and has a moderate anti-secretory effect and beneficial for gastric and duodenal ulcer healing. (Said et al., 1986; Huwez et al., 1986; Al-Habbal et al., 1984). *Samagh-e-arabi* (*Acacia arabica*) contains tannins, saponins, glycosides, phenols terpenoids and flavonoids that can be easily hydrolyzed, this property of *samagh-arabi* has anti-bacterial and anti-inflammatory attributes. (Sultana et al., 2007). It has been reported that licorice extract showed potent eradication effect against *H. pylori* strain (Kakka et al., 1998; Krausse et al., 2004; Marjan et al., 2013). Other researchers found anti *H. pylori* activity of *Plantago ovate* (Nabati et al., 2012). Furthermore Omayma K.H. and M. Amin et al conducted in vitro studies and found anti-bacterial and anti-*H. Pylori* activity of *acacia arabica*. (Nabati et al., 2012; Omayma et al; 2011) *Mastagi* showed anti- *H. Pylori* effect not only in vitro but in clinical trial as well. (Ghani et al., 2005; Farhad et al., 1998). In 1999 ,Gharzouli K et al found gastro-protective effect of tannic acid and the aqueous extract of from *Quercus ilex* L. root bark, *Punica granatum* L. fruit peel and *Artemisia herba-alba* Asso leaves in rats against ethanol-induced gastric damage and suggested that monomeric and polymeric polyphenols can strengthen the gastric mucosal barrier. (Gharzouli et al., 1999) On the other hand, In 2005 Ajai Kumar K.B., et al conducted in vivo research on the inhibition of gastric mucosal injury by *Punica granatum* methanolic extract and they revealed the gastroprotective effect of the extract through antioxidant mechanism. (Ajai et al., 2005) Additionally Jamal et al suggested that *Tabasheer* and other unani mufrad advia/ single drugs are safe and cost effective in gastric ulceration. (Jamal et al, 2006) In 2010, an experimental study conducted on Anti-ulcer effect of hydroalcoholic extract of *Tukhme Kishneez* (*Coriandrum sativum* Linn.) in stress induced gastric ulceration in albino rats with Ranitidine as the standard drug and demonstrated that *Tukhm Kishneez* possesses anti-ulcer effect against stress induced gastric ulcer. (Shagufta et al., 2010) Nearly, all Unani physicians have contributed to awareness on the health and disease of Meda/stomach.

CONCLUSION

This review concluded that *Helicobacter* infection affects more than 50% population world widely. The infection of *Helicobacteriosis* is mostly asymptomatic which make the infection chronic due to which diagnosis become too difficult. *Helicobacter* having a zoonotic potentials also and having various other species such as *H. canis*, *H. suis*, *H. pullorum* which are transmitted from animals and foods of animal origin (meat and milk). In this era, in where antibiotic resistance is more, we can overcome this situation with the help of Unani drugs.

CONFLICT OF INTEREST

The authors declared no conflict of Interest

REFERENCES

- [1] Aziz, “Makhzan-e-Sulemani”, Munshi Nawal Kishore, Lucknow. 1301 A.H., 305-306.
- [2] A.H. Jurjani, “Zakheera Khaearzam Shahi” (Translated by Hkm Hussain Khan), (1903) Matba Nami Nawal Kishore, Lucknow, vol. I, pp. 367-368.
- [3] S. B. S. Qurrah. “Azzakhirah Fit Tib (Makhtootah)”. vol.3: pp.24
- [4] Z. A. Marwan. “Kitabut Taiseer (Urdu translation)”. Central Council for Research in Unani Medicine, New Delhi. Pp. 125-126
- [5] Jamal, A. Siddiqui, Tajuddin and M. A. Jafri. “A review on gastric ulcer remedies used in Unani System of Medicine”. Natural Product Radiance. 2006; Vol. 5(2): pp.153-159.
- [6] Mohd, J. Zoobi, “Evaluation of Antiulcer Activity of Punica granatum Flower Extract in Experimental Animals. International journal of Research in Ayurveda and Pharmacy. 2011; Vol.2 (4): pp. 210-213.
- [7] S. Kakka, N. Shankar, R.S. Ashok, K.K. Saxena, S. Lala and V.K. Shivastava. “Effect of water soluble portion of alcoholic extract of root of *G. glabra* Linn on acute inflammation.” Indian Journal of pharmacology. 1998; vol. 30: pp. 117.
- [8] C.P. Khare. “Encyclopedia of Indian medicinal plants”. New York: Springer-Verlag; 2004; pp. 233-235.
- [9] Rahida Hilal. “Safety And Efficacy of Unani Compound Drug in Helicobacter Pylori Positive Antral Gastritis (Warm-E-Meda Patients): A Controlled Study.” IOSR Journal of Dental and Medical Sciences (IOSR-JDMS) 16.9 (2017): 81-88
- [10] M. S. Al-Said, A.M. Ageel, and N.S. Parmer. “Evaluation of mastic, a crude drug obtained from *Pistacia lenticus* for gastric and duodenal ulcer activity”. Ethnopharmacol. 1986; vol. 15: pp. 271-278.
- [11] F. U. Huwez and M.J. Al-Habbal. “Mastic in treatment of benign gastric ulcers”. Gastroenterol. Jpn. 1986; vol. 21: pp. 273-274.
- [12] M. J. Al-Habbal, Z. Al-Habbal and F.U. Huwez. “A double-blind controlled clinical trial on mastic and placebo in the

- treatment of duodenal ulcer". J. Clin. Exp. Pharm. Physiol. 1984; vol. 11: pp. 541-544.
- [13] Sultana, F. Anwar and R. Przybylski. "Antioxident activity of phenolic components presents in bark of *Azadirachta indica*, *Terminalia arjuna*, *Acacia nilotica* and *Eugenia jambolana* Lam. Trees". Food Chem. 2007; vol. 104: pp. 1106-1114.
- [14] R. Krause, J. Bielenberg, W. Blackchek, U. Ullmann. "In Vitro anti-*Helicobacter pylori* activity of extractum liquoritial, glycyrrhizus and its metabolites", J. Antimicrob. chemother. 2004; vol. 54(1): pp. 243-246.
- [15] R. Marjan, M. Davood, J. Sara, E. Majid and S.F. Mehdi, "The Healing Effect of Licorice (*Glycyrrhiza glabra*) on *Helicobacter pylori* infected peptic ulcers." J Res Med Sci. 2013; vol. 18(6): pp. 532-533.
- [16] Castillo-Juarez, V. Gonzalez, H. Jaime-Aguilar, G. Martinez and E. Linares, et al, "Anti-*Helicobacter pylori* activity of plants used in Maxican traditional medicine for gastrointestinal disorders.", J. Ethnopharmacol. vol. 122: pp.402-405.
- [17] F. Nabati, M. Mojab, M. Hbib-Rezae, K. Bagherzadeh, M. Amanlou and B. Yousefi, "Large scale screening of commonly used Iranian traditional medicinal plants against urease activity." DARU J. Pharm. Sci. 2012; vol. 2. 10.1186/2008-2231-20-72.
- [18] K. H. Omayma, M.Y. Mohamed, E. Mona. "Possible protective effect of gum Arabic on experimentally induced gastric ulcer in adult male albino rats: A histological and immunohistochemical study." Egypt. J. Histol. 2011; vol. 34: pp. 546-553.
- [19] Ghani. Khazain-al-advia. Vol. 1-4, Idara Kitab-ul-shifa, Darya ganj, New Delhi. 2005. SH offset press, New Delhi, India.
- [20] U.H. Farhad and T. Debbie and A. A. D. Ala-Aldeen "Mastic gum kills *Helicobacter pylori*". N Engl. J. Med. 1998: 339:1946/ Dec. 24, 1998/ DOI.10.1056/NEJM 199812243392618.
- [21] Gharzouli K, Khennouf S, Amira S, Gharzouli A. Effects of aqueous extracts from *Quercus ilex* L. root bark, *Punica granatum* L. fruit peel and *Artemisia herba-alba* Asso. leaves on ethanol-induced gastric damage in rats. Phytoter. Res. 1999; vol. 13(1): pp. 42-5.
- [22] Ajai kumar KB, Asheef M, Babu BH, Padikkala J. The inhibition of gastric mucosal injury by *Punicagranatum* L. (pomegranate) methanolic extract, J Ethnopharmacol. 2005; vol. 96(1-2): pp.171-6.
- [23] Shagufta Nikhat, Ghufuran Ahmad, Nasreen Jahan. Anti-ulcer effect of Tukhme Kishneez (*Coriandrum sativum* Linn.) in stress induced gastric ulceration in albino rats, Unani Medicus. 2010; vol. 1 (1): 62-70.
- [24] Farinha, P. and Gascoyne, R. D. 2005. *Helicobacter pylori* and MALT lymphoma. *Gastroenterol.* 128: 1579-1605.
- [25] Amieva, M. R. and El-Omar, E. M. 2008. Host-bacterial interactions in *Helicobacter pylori* infection. *Gastroenterology.* 134:306-323.
- [26] Suerbaum, S. and Josenhans, C. 2007. "*Helicobacter pylori* evolution and phenotypic diversification in a changing host," *Nature Rev. Microbiol.* 5(6): 441-452.
- [27] Lochhead, P. and El-Omar, E. M. 2007. *Helicobacter pylori* infection and gastric cancer. *Best Pract. Res. Clin. Gastroenterol.* 21: 281-297.
- [28] Kusters, J. G., van Vliet, A. H. and Kuipers, E. J. 2006. Pathogenesis of *Helicobacter pylori* *Infect. Clin. Microbiol. Rev.* 19: 449-490.
- [29] Correa, P. 2004. The biological model of gastric carcinogenesis. *IARC Sci. Publ.* 301-310.
- [30] Gonzalez, C. A., Lujan-Barroso, L., Bueno-de-Mesquita, H. B., Jenab, M., Duell, E. J. and Agudo, A. 2012. Fruit and vegetable intake and the risk of gastric adenocarcinoma: a reanalysis of the European Prospective Investigation into Cancer and Nutrition (EPIC-EURGAST) study after a longer follow-up. *Int. J. of Cancer.* 131: 2910- 2919.
- [31] Wadhwa, R., Song, S., Lee, J. S., Yao, Y., Wei, Q. and Ajani, J. A. 2013. Gastric cancer- molecular and clinical dimensions. *Nature Reviews. Clin. Oncology.* 10: 643- 655.
- [32] Globocan. 2012. Available at: <http://globocan.iarc.fr>. Accessed on June 22, 2017.
- [33] Chmiela, M., Karwowska, Z., Gonciarz, W., Allushi, B. and Stączek, P. 2017. Host pathogen interactions in *Helicobacter pylori* related gastric cancer. *World J. Gastroenterol.* 23(9): 1521-1540.
- [34] Torre, L. A., Bray, F., Siegel, R. L., Ferlay, J., Lortet-Tieulent, J. and Jemal, A. 2015. Global cancer statistics, 2012. *CA: A Cancer Journal for Clinicians.* 65: 87-108.
- [35] Parsonnet, J. 1994. Gastric adenocarcinoma and *Helicobacter pylori* infection. *West. J. Med.* 161: 60.
- [36] Farinha, P. and Gascoyne, R. D. 2005. *Helicobacter pylori* and MALT lymphoma. *Gastroenterol.* 128: 1579-1605.
- [37] Patel, A., Shah, N. and Prajapati, J. B. 2014. Clinical application of probiotics in the treatment of *Helicobacter pylori* infection da brief review. *J. Microbiol. Immunol. Infect.* 47(5): 429-437.
- [38] Suerbaum, S. and Josenhans, C. 2007. "*Helicobacter pylori* evolution and phenotypic diversification in a changing host," *Nature Rev. Microbiol.* 5(6): 441-452.
- [39] Stolte, M., Bayerdorffer, E., Morgner, A., Alpen, B., Wundisch, T., Thiede, C. and Neubauer, 2002. *Helicobacter* and gastric MALT lymphoma. *Gut.* 50(3): 19-24.
- [40] Wotherspoon, A. C., Ortiz-Hidalgo, C., Falzon, M. R. and Isaacson, P. G. 1991. *Helicobacter pylori* associated gastritis and primary B-cell gastric lymphoma. *Lancet.* 338: 1175-1176.
- [41] Mbulaiteye, S. M., Hisada, M. and El-Omar, E. M. 2009. *Helicobacter Pylori* associated global gastric cancer burden. *Front. Biosci. (Landmark Ed).* 14: 1490-1504.
- [42] Kawakubo, M., Horiuchi, K. and Matsumoto, T. 2018. Cholesterol- α -glucosyltransferase gene is present in most *Helicobacter* species including gastric non-*Helicobacter pylori* helicobacters obtained from Japanese patients. *Helicobacter.* 23: e12449.
- [43] Liu, J., He, L., Haesebrouck, F., Gong, Y., Flahou, B., Cao, Q. and Zhang, J. 2014. Prevalence of Coinfection with Gastric Non-*Helicobacter pylori* *Helicobacter* (NHPH) Species in *Helicobacter pylori*

- infected Patients Suffering from Gastric Disease in Beijing, China. *Helicobacter*. 20: 284–290.
- [44] Van den Bulck, K., Decostere, A., Baele, M., Driessen, A., Debongnie, J. C., Burette, A., Stolte, M., Ducatelle, R. and Haesebrouck, F. 2005. Identification of non- *Helicobacter pylori* spiral organisms in gastric samples from humans, dogs, and cats. *J. Clin. Microbiol.* 43(5): 2256-2260.
- [45] Lavelle, J. P., Landas, S., Mitros, F.A. and Conklin, J. L. 1994. Acute gastritis associated with spiral organisms from cats. *Dig. Dis. Sci.* 39: 744-750.
- [46] Yoshimura, M., Isomoto, H., Shikuwa, S., Osabe, M., Matsunaga, K., Omagari, K., Mizuta, Y., Murase, K., Murata, I. and Kohno, S. 2002. A case of acute gastric mucosal lesions associated with *Helicobacter heilmannii* infection. *Helicobacter*. 7: 322- 326.
- [47] Haesebrouck, F., Pasmans, F., Flahou, B., Chiers, K., Baele, F. T., Fteyns, T., Decostere, A. and Ducatelle, R. 2009. Gastric *Helicobacters* in domestic animals and non- human primates and their significance for human health. *Clin. Microbiol. Rev.* 22: 202–223.
- [48] Debongnie, J. C., Donnay, M., Mairesse, J., Lamy, V., Dekoninck, X. and Ramdani, B. 1998. Gastric ulcers and *Helicobacter heilmannii*. *Eur. J. Gastroenterol. Hepatol.* 10: 251-254.
- [49] Seo, W.J., Park, C. S., Cho, Y.J., Cha, K. W., Lee, S. W., Lim, S. T., Sung, Y.H. and Baek, R. 2003. A case of gastric ulcer induced by *Helicobacter heilmannii*-like organism. *Korean J. Gastroenterol.* 42: 63-66.
- [50] Jhala, D., Jhala, N., Lechago, J. and Haber, M. 1999. *Helicobacter heilmannii* gastritis: association with acid peptic diseases and comparison with *Helicobacter pylori* gastritis. *Mod. Pathol.* 12: 534-538.
- [51] Iwanczak, B., Biernat, M., Iwanczak, F., Grabinska, J., Matusiewicz, K. and Gosciniak, G. 2012. The clinical aspects of *Helicobacter heilmannii* infection in children with dyspeptic symptoms. *J. Physiol. Pharmacol.* 63: 133-136.
- [52] Swennes, A. G., Parry, N. M. A. and Feng, Y. 2016. Enterohepatic *Helicobacter* spp. in cats with non-haematopoietic intestinal carcinoma: a survey of 55 cases. *J. Med. Microbiol.* 65: 814820.
- [53] Joosten, M., Linden, S., Rossi, M., Tay, A.C.Y., Skoog, E. and Medea, P. 2015. Divergence between the highly virulent *Helicobacter heilmannii* and its closest relative, the low-virulence *Helicobacter ailurogastricus* sp. nov. *Infect Immun.* 84: 293–306.
- [54] Mazzucchelli, L., Wilder-Smith, C. H., Ruchti, C., Meyer-Wyss, B. and Merki, H. S. 1993. *Gastrospirillum hominis* in asymptomatic, healthy individuals. *Dig. Dis. Sci.* 38: 2087-2089.
- [55] Morgner, A., Lehn, N., Andersen, L. P., Thiede, C., Bennedsen, M., Trebesius, K., Neubauer, B., Neubauer, A., Stolte, M. and Bayerdorffer, E. 2000. *Helicobacter heilmannii*- associated primary gastric low-grade MALT lymphoma: complete remission after curing the infection. *Gastroenterol.* 118: 821-828.
- [56] Thomas-Marques, L., Yaziji, N., Bouche, O., Diebold, M. D., Cadot, G. and Thieffn, G. 2005. *Helicobacter heilmannii*-associated low-grade gastric MALT lymphoma: a new case of complete remission after eradication. *Gastroenterol. Clin. Biol.* 29: 476-477.
- [57] Dieterich, C., Wiesel, P., Neiger, R., Blum, A. and Cortesy-Theulaz, I. 1998. Presence of multiple “*Helicobacter heilmannii*” strains in an individual suffering from ulcers and in his two cats. *J. Clin. Microbiol.* 36: 1366- 1370.
- [58] Kaklikkaya, N., Ozgur, O., Aydin, F. and Cobanoglu, U. 2002. *Helicobacter heilmannii* as causative agent of chronic active gastritis. *Scand. J. Infect. Dis.* 34: 768-770.
- [59] Asl, A. S., Jamshidi, S., Mohammadi, M., Soroush, M. H., Bahadori, A. and Oghalaie, A. 2010. Detection of atypical cultivable canine gastric *Helicobacter* strain and its biochemical and morphological characters in naturally infected dogs. *Zoonoses Pub Health.* 57: 244-248.
- [60] Hristova, I. M., Grekova, O. and Patel, A. 2017. Zoonotic potential of *Helicobacter* spp. *J. of Microbiol. Immunol. and Infect.* 50: 265-269.
- [61] Seim-Wikse, T., Jorundsson, E., Nodtvedt, A., Grotmol, T., Bjornvad, C. R. and Kristensen, T., 2013. Breed predisposition to canine gastric carcinoma – a study based on the Norwegian canine cancer register. *Acta Veterinaria Scandinavica.* 55: 25.
- [62] De Groote, D., Van Doorn, L. J. and Van den Bulck, K. 2005. Detection of non-*pylori* *Helicobacter* species in “*Helicobacter heilmannii*”- infected humans. *Helicobacter.* 10: 398-406.
- [63] Flahou, B., Van Deun, K., Pasmans, F., Smet, A., Volf, J., Rychlik, I., Ducatelle, R. and Haesebrouck, F. 2012. The local immune response of mice after *Helicobacter suis* infection: strain differences and distinction with *Helicobacter pylori*. *Vet. Res.* 43:75.
- [64] Matsui, H., Takahashi, T. and Murayama, S. Y. 2014. Development of new PCR primers by comparative genomics for the detection of *Helicobacter suis* in gastric biopsy specimens. *Helicobacter.* 19:260-271.
- [65] Kumar, S., Haesebrouck, F., Pasmans, F., Flahou, B., Dewulf, J., Chiers, K. and Ducatelle, R. 2010. An experimental *Helicobacter suis* infection reduces daily weight gain in pigs. *Proc IPVS. Vancouver, Canada.* 080:117.
- [66] De Bruyne, E., Flahou, B., Chiers, K., Meyns, T., Kumar, S., Vermoote, M., Pasmans, F., Millet, S., Dewulf, J., Haesebrouck, F. and Ducatelle, R. 2012. An experimental *Helicobacter suis* infection causes gastritis and reduced daily weight gain in pigs. *Vet. Microbiol.* 160: 449–454.
- [67] Bosschem, I., Flahou, B. and Bakker, J. 2016. Comparative virulence of in viro cultures primate- and pig-associated *Helicobacter suis* strains in a BALB/c mouse and Mongolian gerbil model. *Helicobacter.* DOI:10.1111/hel12349.
- [68] Blaecher, C., Bauwens, E., Tay, A., Peters, F., Dobbs, S., Dobbs, J., Charlett, A., Ducatelle, R., Haesebrouck, F. and Smet, A. 2017. A novel isolation protocol and probe- based RT-PCR for diagnosis of gastric infections with the zoonotic pathogen *Helicobacter suis*. *Helicobacter.* 22: e12369.
- [69] Blaecher, C., Smet, A. and Flahou, B. 2013. Significantly high frequency of *Helicobacter suis* in patients with idiopathic parkinsonism in contrast to control patients. *Aliment Pharmacol Ther.* 38(11-12):1347–1353.
- [70] Overby, A., Murayama, S. Y. and Michimae, H. 2017. Prevalence of gastric non-*Helicobacter pylori* helicobacters in Japanese

- patients with gastric disease. *Digestion*. 95: 61-66.
- [71] Ceelen, L., Decostere, A., Martel, A., Pasmans, F. and Haesebrouck, F. 2006. First report of *Helicobacter pullorum* in the faeces of a diarrhoeic psittacine bird (*Psephotus haematogaster*). *Vet Res*. 159:389-390.
- [72] Stanley, J., Linton, D., Burnens, A.P., Dewhirst, F. E., On, S.L.W., Porter, A., Owen, R.J. and Costas, M. 1994. *Helicobacter pullorum* sp. nov.- genotype and phenotype of a new species isolated from poultry and from human patients with gastroenteritis. *Microbiol*. 140: 3441-3449.
- [73] Turk, M.L., Cacioppo, L. D. and Ge, Z. 2012. Persistent *Helicobacter pullorum* colonization in C57BL/6NTac mice: a new mouse model for an emerging zoonosis. *J. Med. Microbiol*. 61: 720-728.
- [74] Kamboj, A. K., Cotter, T. G. and Oxentenko, A. S. 2017. *Helicobacter pylori*: the past, present, and future in management. *Mayo Clin. Proc*. 92:599-604.
- [75] Guarner, J., Kalach, N., Elitsur, Y. and Koletzko, S. 2010. *Helicobacter pylori* diagnostic tests in children: Review of the literature from 1999 to 2009. *Eur. J. Pediatr*. 169(1):15-25.
- [76] McColm, A. A., Bagshaw, J. A. and O'Malley, C. F. 1993. Development of a ¹⁴C-urea breath test in ferrets colonized with *Helicobacter mustelae*: effects of treatment with bismuth, antibiotics, and urease inhibitor. *Gut*. 34: 181-186.
- [77] Genta, R. M., Robason, G. O. and Graham, D. Y. 1994. Simultaneous visualization of *Helicobacter pylori* and gastric morphology: a new stain. *Hum. Pathol. Mar*. 25(3): 221-226.
- [78] Van Duynhoven, Y.T. and Jonge, R.D. 2001. Transmission of *Helicobacter pylori*: a role for food? *Bulletin of the World Health Organization*, 79:455-460.
- [79] Quaglia, N.C., Dambrosio, A., Normanno, G., Parisi, A., Firinu, A., Lorusso, V. and Celano G.V. 2007. Survival of *Helicobacter pylori* in artificially contaminated ultrahigh temperature and pasteurized milk. *Food microbiology*, 24(3):296-300.
- [80] Quaglia, N.C., Dambrosio, A., Normanno, G., Parisi, A., Patrono, R., Ranieri, G., Rella, A. and Celano, G.V., 2008. High occurrence of *Helicobacter pylori* in raw goat, sheep and cow milk inferred by glmM gene: a risk of food-borne infection? *International journal of food microbiology*, 124(1):43-47.
- [81] Dore, M.P., Sepulveda, A.R., El-Zimaity, H., Yamaoka, Y., Osato, M.S., Mototsugu, K., Nieddu, A.M., Realdi, G. and Graham, D.Y., 2001. Isolation of *Helicobacter pylori* from sheep—implications for transmission to humans. *The American Journal of Gastroenterology*, 96(5):1396.